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Phosphorus, Sulfur, and Silicon and the Related Elements

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713618290>

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To cite this Article Głowacki, Zdzisław and Hoffmann, Maria(1991) ³¹P NMR NON-EQUIVALENCE OF THE DIASTEREOISOMERIC PHOSPHONODIDEPSIPEPTIDES. PART II¹, Phosphorus, Sulfur, and Silicon and the Related Elements, 63: 1, 171 – 175

To link to this Article: DOI: 10.1080/10426509108029440

URL: <http://dx.doi.org/10.1080/10426509108029440>

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³¹P NMR NON-EQUIVALENCE OF THE DIASTEREOISOMERIC PHOSPHONODIDEPSIPEPTIDES. PART II¹

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(Received April 16, 1991; in final form May 13, 1991)

³¹P chemical shifts are reported for 24 different models of diastereoisomeric phosphonodidepsipeptides coupled from the N-protected L-aminoacids and dibenzyl esters of chiral 1-hydroxyalkylphosphonic acids. The ³¹P NMR nonequivalence variations with the changing of dipeptide structure, solvent and temperature were investigated. Amide *cis-trans* isomerism were observed in the case of Z-L-proline derived dipeptides.

Key words: Enantiomeric excess; 1-hydroxyalkylphosphonic acids; ³¹P magnetic nonequivalence; phosphonodidepsipeptides.

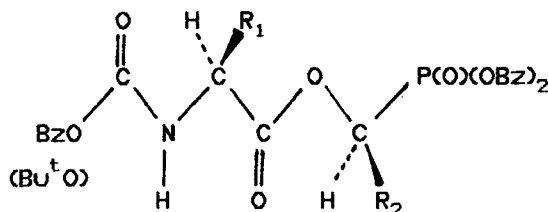
INTRODUCTION

In the first part¹ we presented convenient methods for enantiomeric composition determination of chiral 1-hydroxyalkylphosphonic acids from the ³¹P NMR spectra of their diastereoisomeric phosphonodidepsipeptides, derivatives of the N-protected natural aminoacids. In order to study correlations between the structure of the phosphonodidepsipeptides and the diastereotopic chemical shift nonequivalences, more than 20 additional phosphonodidepsipeptides were synthesized and their ³¹P NMR spectra were recorded in different conditions.

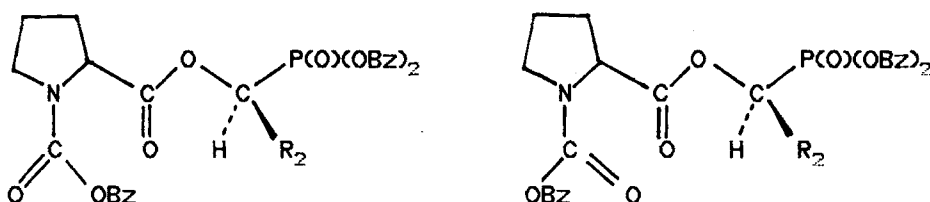
NMR spectra of aminoacids, peptides, their derivatives^{2–5} and phosphonic analogs^{1,6,7} are frequently used to measure enantiomeric composition, to follow racemization during coupling reaction or to follow resolution processes, also to control diastereoisomeric purity of the resulting product.

RESULTS AND DISCUSSION

The pairs of 1-hydroxyalkylphosphonic acid ester and N-protected aminoacid which were coupled by means of the DCC method and the ³¹P NMR data of the resulting phosphonodidepsipeptides (**1–24**, depicted in Scheme I), L(+) or L(–), are collected in Table I. ³¹P NMR spectra showed the persistence of two distinguishable diastereoisomers when the racemate samples of chiral 1-hydroxyalkylphosphonic acid esters were used. NMR spectra of Z-L-proline derivatives **24** are more complex than those of other aminoacid dipeptides, the latter showed signals of only the transoid form (Z conformer).^{9,10} Amide *cis-trans* isomerism has been the subject of a number of studies, especially that of proline residue, in dipeptides, polypeptide and in protein structures.^{10–12} Both, cisoid and transoid forms, are observed in the



Transoid form of didepsipeptides 1 - 23.



Cisoid and transoid forms of proline didepsipeptides 24.

SCHEME I

^{31}P NMR spectra of **24**. The coalescence of the two peaks was recorded near 330 K in CDCl_3 solution. The difference between $\delta^{31}\text{P}$ of *cis* and *trans* forms depends on didepsipeptide configuration, and is larger for L(−) isomer (0.144 ppm and 0.053 ppm of L(−) and L(+), respectively). Higher sample temperature destruct the defined conformational preference and reduce the differences between *cis* and *trans* conformer as well as magnetic nonequivalence of diastereoisomers. The same effects are observed when the H-bonded structure of didepsipeptide is destroyed by polar solvents such as pyridine or DMSO (1 and 2).

The variations of $\Delta \delta^{31}\text{P}$ with the substituent at the hydroxyphosphonates asymmetric centre and the N-protected L-aminoacids, are collected in Table II. The best chiral derivatizing agent is Boc-N-protected phenylalanine ($\Delta \delta^{31}\text{P} = 0.50 - 0.20$ ppm). Assuming, the similar conformer population of the didepsipeptide reported in the table, the observed $\delta^{31}\text{P}$ anisochrony is related only to R_2 . Highest $\Delta \delta^{31}\text{P}$ values were recorded for isopropyl and isobutyl at the chiral centre, surprisingly the anisotropic phenyl revealed smallest magnetic differentiation. The protecting groups (Z- or Boc-) do not change the magnitude of nonequivalence.

Only for two of the here reported hydroxyphosphonates the absolute configurations have been determined; S(−) configuration of dimethyl α -hydroxybenzylphosphonate¹³ and R(−) of dibenzyl 1-hydroxyisopentylphosphonate.¹⁴ ^{31}P NMR signals of L-aminoacids and R-hydroxybenzyl- and R-hydroxyisopentyl-phosphonates didepsipeptides are always shifted to lower field (2, 5, 12, 21 and 24).

TABLE I
³¹P NMR chemical shifts of the phosphonate groups in the
 diastereoisomeric phosphonodidepsipeptides

PDP ¹	N-L-AA	R ₁	R ₂	$\delta(^{31}\text{P})$		$\Delta\delta$ [ppm]	Notes
				L(+)	L(-)		
1	Boc-L-Phe,	Bz	(+)i-Pr rac	20.408			a*, CDCl ₃
				20.434	20.866	0.432	a*, CDCl ₃
				21.022	21.514	0.492	CDCl ₃
				21.096	21.582	0.486	b, CDCl ₃
				21.461	22.049	0.588	benzene
2			S(+)i-Bu R(-) rac	21.401	21.785	0.384	pyridine
				21.867			CDCl ₃
					22.279		CDCl ₃
				21.865	22.280	0.415	CDCl ₃
				22.514	22.929	0.415	benzene
3			rac n-Pr	22.421	22.719	0.298	pyridine
				22.103	22.411	0.308	DMSO
4			rac Me	22.108			
				21.693		0.415	CDCl ₃
5			R(+)Ph rac	22.492			
				22.103		0.389	CDCl ₃
				22.818			
				22.446		0.372	b, benzene
6	Z-L-Phe	Bz	rac i-Pr	18.505			a*, CDCl ₃
				18.517	18.318		a*, CDCl ₃
				21.563			
				21.129		0.434	CDCl ₃
				22.355			
7			rac i-Bu	21.941		0.414	CDCl ₃
				21.974			
8			rac n-Pr	21.559		0.415	CDCl ₃
				22.379			
9			rac Me	21.989		0.390	b, CDCl ₃
				18.723			
10			rac Ph	18.527		0.196	CDCl ₃
				21.249			
11	Boc-L-Val	i-Pr	(+)i-Pr	21.246	21.665	0.419	CDCl ₃
				22.188	22.528	0.340	CDCl ₃
12			R(-)i-Bu	22.167			
				21.851		0.316	CDCl ₃
13			rac n-Pr	22.522			
				22.315		0.207	CDCl ₃
14			rac Me	22.802			
				22.600		0.202	b, benzene
15			R(+) Ph rac	18.751			CDCl ₃
				18.728		0	CDCl ₃
16	Z-L-Val	i-Pr	rac i-Pr	22.310			
				21.954		0.356	CDCl ₃
17			rac i-Bu	21.634			
				21.218		0.416	CDCl ₃
18	Boc-L-Ile	sec-Bu	rac i-Pr	21.651			
				21.230		0.421	b, CDCl ₃
19			rac i-Bu	22.555			
				22.247		0.308	b, CDCl ₃
20			rac Ph	18.738		0	b, CDCl ₃
				18.670		0	CDCl ₃
21	Boc-L-Leu	i-Bu	R(-)i-Bu rac		22.347		CDCl ₃
				22.177	22.338	0.161	CDCl ₃

TABLE I (continued)

PDP ¹	N-L-AA	R ₁	R ₂	$\delta(^{31}\text{P})$		$\Delta\delta$ [ppm]	Notes
				L(+)	L(-)		
22			rac Me	22.463		0.103	CDCl_3
				22.360			
				22.808		0.145	b, benzene
				22.663			
23	Z-L-Leu	i-Bu	rac i-Bu	22.284		0.190	CDCl_3
				22.094			
24	Z-L-Pro		R(-) i-Bu	22.727			303°K, CDCl_3
				22.583			
				22.612			
				22.499			
			rac	22.495		0.233	330°K, CDCl_3
				22.459	22.738		
				22.406	22.594		
			rac	22.293	22.468	0.175	330°K, CDCl_3

¹Phosphonodipeptides (PDP) obtained from N-protected L-aminoacids (N-L-AA) and (-), (+) or racemate (rac) of 1-hydroxyalkylphosphonic acid dibenzylesters (HPE) as indicated in the table.

^a*ERRATUM: these data were not correctly rewritten in Table I of the Part I (ref. 1), 1a* recorded on Jeol FX90Q at 36.27 MHz.

^bBruker AC-200 at 80.02 MHz, all other spectra from Bruker MSL-300 at 121.5 MHz.

Boc = t-butoxycarbonyl group, Z = benzyloxycarbonyl group.

TABLE II

The variations of $\Delta \delta^{31}\text{P}$ (ppm) between L(+) and L(-) with the substituent at the hydroxyphosphonates asymmetric centre and the N-protected L-aminoacids

R ₂	N-protected aminoacid		
	Boc-Phe {	Z-Phe $\Delta\delta^{31}\text{P}$	Boc-Val }
i-Pr	0.492	0.434	0.419
i-Bu	0.415	0.414	0.340
n-Pr	0.415	0.415	0.316
Me	0.389	0.390	0.207
Ph	0.199	0.196	0

EXPERIMENTAL

The general procedure for coupling of phosphonodipeptides **1–24** by means of DCC have been reported previously.^{1,8} The ³¹P NMR spectra of the products in CDCl_3 (benzene, pyridine or DMSO) were recorded on a FT-NMR spectrometer Bruker MSL-300 at 121.5 MHz and Bruker AC-200 at 81.02 MHz. An 85% H_3PO_4 solution was used as an external reference. Typical conditions: spectral width 20000 Hz, number of scans 50–100 and digital resolution 1.2 Hz per data point.

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